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(54) Title: H ₂ O ₂ -STABLE PEROXIDASE VARIANTS		
(57) Abstract The present invention relates to novel variants of <u>Coprinus cinereus</u> peroxidase showing excellent hydrogen peroxide stability.		

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H₂O₂-STABLE PEROXIDASE VARIANTS

FIELD OF INVENTION

The present invention relates to a novel variant of peroxidase, and a bleaching or detergent composition comprising the peroxidase variant.

BACKGROUND OF THE INVENTION

The use of bleaching agents in washing procedures and as constituents of detergent compositions is well known in the art. Thus, bleaching agents are incorporated in or sold as constituents of a major part of the commercially available detergent compositions. Important conventional bleaching agents incorporated in detergent compositions are compounds which act as precursors of hydrogen peroxide formed in the course of the washing procedure. Perborates and percarbonates are the most important examples of compounds which are employed as bleaching agents and which exert a bleaching effect in this fashion. The detailed mechanism of bleaching by means of these bleaching agents is not known at present, but it is generally assumed that the hydrogen peroxide formed during washing converts coloured substances (responsible for stains on fabric) into non-coloured materials by oxidation and that some oxidation of the coloured substances may also take place due to their direct interaction with perborate or percarbonate.

It has been found that peroxidases, utilizing hydrogen peroxide as substrate, are able to enhance the bleaching effect of hydrogen peroxide during washing. The use of peroxidases for bleaching stains on fabrics is described in WO 89/09813. It was also found that coloured substances leached from dyed fabrics could be bleached by means of peroxidases together with hydrogen peroxide. The use of peroxidases for inhibiting dye transfer in this way is described in WO 91/05839.

SUMMARY OF THE INVENTION

It has surprisingly been found that peroxidase variants with an improved stability towards hydrogen peroxide at alkaline conditions may be prepared by recombinant DNA techniques.

Accordingly, the present invention relates to a peroxidase variant with improved hydrogen peroxide stability at alkaline conditions, characterized by insertion, deletion or substitution of one or more amino acid residues in the region from amino acid residue 48 to 56, 76, 109, 214, 239, 258 to 262, 264, 266 to 272 of the parent peroxidase, a Coprinus cinereus peroxidase encoded by the sequence shown in SEQ ID 1.

Information about the crystal structure of the parent peroxidase was obtained by X-ray diffraction, and also by aligning the amino acid sequence of the parent peroxidase to amino acid sequences of other known peroxidases (K.G. Welinder et al., "Structure and evolution of peroxidases" in Plant Peroxidase Biochemistry and Physiology, K.G. Welinder et al. (eds.), University of Copenhagen and Geneva 1993).

In the present context the term "improved hydrogen peroxide stability" is intended to indicate that the peroxidase variant is at least 10% more stable than the parent peroxidase in the presence of hydrogen peroxide at a concentration of up to 20 mM H_2O_2 . More specifically this is intended to mean that the peroxidase variant at one or more temperatures in the temperature interval of from 10°C to 60°C and at one or more H_2O_2 concentrations of up to 20 mM H_2O_2 has an at least 10% longer half-life than the wild type peroxidase (parent peroxidase). (Half-life is determined by fitting residual activities to 1st order decay). The term "alkaline conditions" is intended to indicate the pH-range from 7 to 11.

In other aspects, the present invention relates to a bleaching composition comprising a peroxidase variant according to the invention and a hydrogen peroxide source, optionally additionally comprising an oxidizable substrate; and to a detergent composition comprising a surfactant, a peroxidase

variant according to the invention and a hydrogen peroxide source, optionally additionally comprising an oxidizable substrate.

BRIEF DESCRIPTION OF DRAWINGS

5 The present invention is further illustrated by reference to the accompanying drawing, in which

Fig. 1 shows plasmid pHD414, which is a derivative of the plasmid p775 (described in EP 238023). Plasmid pHD414 is obtained according to Ex.1.

10 DETAILED DISCLOSURE OF THE INVENTION

In the present description and claims, the following abbreviations are used:

Amino Acids:

A	=	Ala	=	Alanine
15V	=	Val	=	Valine
L	=	Leu	=	Leucine
I	=	Ile	=	Isoleucine
P	=	Pro	=	Proline
F	=	Phe	=	Phenylalanine
20W	=	Trp	=	Tryptophan
M	=	Met	=	Methionine
G	=	Gly	=	Glycine
S	=	Ser	=	Serine
T	=	Thr	=	Threonine
25C	=	Cys	=	Cysteine
Y	=	Tyr	=	Tyrosine
N	=	Asn	=	Asparagine
Q	=	Gln	=	Glutamine
D	=	Asp	=	Aspartic Acid
30E	=	Glu	=	Glutamic Acid
K	=	Lys	=	Lysine

R = Arg = Arginine
H = His = Histidine

In describing peroxidase variants according to the invention, the following nomenclature is used for ease of reference:

Original amino acid:position:substituted amino acid(s)

According to this nomenclature, for instance the substitution of lysine by serine in position 48 is as shown:

K48S

10a deletion of lysine in the same position is shown as:

K48*

and insertion of an additional amino acid residue such as tyrosine is shown as:

K48KY

15 Multiple substitutions are separated by pluses, i.e.:

E214L + E239L

representing mutations in positions 214 and 239 substituting leucine for glutamic acid.

The parent peroxidase is encoded by the amino acid
20sequence shown in SEQ ID No. 1. Said sequence is derivable from Coprinus cinereus.

In one embodiment of the peroxidase variant of the invention, one or more amino acid residues are deleted, inserted or substituted in the region from amino acid residue
2548 to 56, 76, 109, 214, 239, 258 to 262, 264, 266 to 272 of the parent peroxidase encoded by the amino acid sequence shown in SEQ ID No. 1.

In other embodiments of the peroxidase variant according to the invention, one or more amino acid residues may
30suitably be substituted as follows

K48S,

V53K,A,

G72Q,

A91C,

35 N92K,

H109K,
Q118E,
M125A, P, W, G, S, T, C, Y, N, D, E, K, R, H,
in particular M125S, T,
5 S147Q,
I152C,
P155C,
M166A, P, W, G, S, T, C, Y, N, D, E, K, R, H,
in particular M166S, T,
10 N192K,
I195K,
V206R,
E214L,
K218R,
15 F229G,
A230C,
E239A, V, L, I, P, F, W, M, G, S, T, C, Y, N, Q, D, K, R, H,
in particular E239K, G, S, L, Q, M,
M242A, P, W, G, S, T, C, Y, N, D, E, K, R, H,
20 in particular M242S, T,
S244C,
S252P,
W258F, H,
M261A, V, L, I, P, F, W, G, S, T, C, Y, N, Q, D, E, K, R, H,
25 in particular M261F, V, I, L, Q,
M268A, V, L, I, P, F, W, G, S, T, C, Y, N, Q, D, E, K, R, H,
in particular M268F, V, I, L, Q,
Y272A, V, L, I, P, F, W, G, S, T, C, M, N, Q, D, E, K, R, H,
in particular Y272F,
30 M276A, P, W, G, S, T, C, Y, N, D, E, K, R, H,
in particular M276S, T,
K278R,
M279A, P, W, G, S, T, C, Y, N, D, E, K, R, H,
in particular M279S, T,
35 A304E,
V314P.

In an alternative embodiment, the peroxidase variant according to the invention may be a fragment of the peroxidase variant described above.

According to the invention, two or more amino acid residues of the peroxidase sequence may also be substituted as follows

10 K41R + K48R,
 V53K + Q118E,
 V53A + E239G,
 I152C + A91C,
 P155C + A230C,
 M166F + E239K,
 G167N + V176L,
 E214L + E239L,
15 R241E + E239K,
 S244C + P155C,
 E239K + M242I + Y272F,
 M166F + E239K + M242I + Y272F,
 M125L + M166F + E239K + M242I + Y272F.

20 According to the invention the substitutions, insertions or deletions disclosed in this invention may be combined with the substitutions, insertions or deletions disclosed in WO 93/24618, in which one or more amino acid residues are deleted, inserted or substituted in the region of
25 from amino acid residue 79 to 94, 125, 153 to 157, 161 to 204, 242, 276 or 279 of the parent peroxidase shown in the present invention as SEQ ID No.1. An example of such a combination is the variant M242I + Y272F + E239K, which has a very good hydrogen peroxide stability as demonstrated in Example 4.

30 The DNA sequence encoding a parent peroxidase may be isolated from any microorganism producing the peroxidase in question by various methods well known in the art. First a genomic DNA and/or cDNA library should be constructed using chromosomal DNA or messenger RNA from the organism that
35 produces the peroxidase to be studied. Then, if the amino acid sequence of the peroxidase is known, homologous, labelled

oligonucleotide probes may be synthesized and used to identify peroxidase-encoding clones from a genomic library of bacterial DNA, or from a fungal cDNA library. Alternatively, a labelled oligonucleotide probe containing sequences homologous to speroxidase from another strain of fungus could be used as a probe to identify peroxidase-encoding clones, using hybridization and washing conditions of lower stringency.

Another method for identifying peroxidase-producing clones involves inserting fragments of genomic DNA into an expression vector, such as a plasmid, transforming peroxidase-negative bacteria with the resulting genomic DNA library, and then plating the transformed bacteria onto agar containing a substrate for peroxidase. Those bacteria containing peroxidase-bearing plasmid will upon addition of hydrogen peroxide produce colonies surrounded by a halo of clear agar, due to oxidation of the substrate by secreted peroxidase.

Alternatively, the DNA sequence encoding the enzyme may be prepared synthetically by established standard methods, e.g. the phosphoamidite method described by S.L. Beaucage and M.H. Caruthers, Tetrahedron Letters **22**, 1981, pp. 1859-1869, or the method described by Matthes et al., The EMBO J. **3**, 1984, pp. 801-805. According to the phosphoamidite method, oligonucleotides are synthesized, e.g. in an automatic DNA synthesizer, purified, annealed, ligated and cloned in appropriate vectors.

Finally, the DNA sequence may be of mixed genomic and synthetic, mixed synthetic and cDNA or mixed genomic and cDNA origin prepared by ligating fragments of synthetic, genomic or cDNA origin (as appropriate), the fragments corresponding to various parts of the entire DNA sequence, in accordance with standard techniques. The DNA sequence may also be prepared by polymerase chain reaction (PCR) using specific primers, for instance as described in US 4,683,202 or R.K. Saiki et al., Science **239**, 1988, pp. 487-491.

35 Once a peroxidase-encoding DNA sequence has been isolated, and desirable sites for mutation identified, mutations may be introduced using synthetic oligonucleotides. These

oligonucleotides contain nucleotide sequences flanking the desired mutation sites; mutant nucleotides are inserted during oligonucleotide synthesis. In a specific method, a single-stranded gap of DNA, bridging the peroxidase-encoding sequence, is created in a vector carrying the peroxidase gene. Then the synthetic nucleotide, bearing the desired mutation, is annealed to a homologous portion of the single-stranded DNA. The remaining gap is then filled in with DNA polymerase I (Klenow fragment) and the construct is ligated using T4 ligase. A specific example of this method is described in Morinaga et al., (1984, *Biotechnology* 2:646-639). U.S. Patent number 4,760,025, by Estell et al., issued July 26, 1988, discloses the introduction of oligonucleotides encoding multiple mutations by performing minor alterations of the cassette, however, an even greater variety of mutations can be introduced at any one time by the Morinaga method, because a multitude of oligonucleotides, of various lengths, can be introduced.

Another method of introducing mutations into peroxidase-encoding sequences is described in Nelson and Long, *Analytical Biochemistry* 180, 1989, pp. 147-151. It involves the 3-step generation of a PCR fragment containing the desired mutation introduced by using a chemically synthesized DNA strand as one of the primers in the PCR reactions. From the PCR-generated fragment, a DNA fragment carrying the mutation may be isolated by cleavage with restriction endonucleases and reinserted into an expression plasmid.

According to the invention, a mutated peroxidase-coding sequence produced by one of the methods described above, or any alternative methods known in the art, can be expressed, in enzyme form, using an expression vector which typically includes control sequences encoding a promoter, operator, ribosome binding site, translation initiation signal, and, optionally, a repressor gene or various activator genes. To permit the secretion of the expressed protein, nucleotides encoding a signal sequence may be inserted prior to the peroxidase-coding sequence. For expression under the direction of control sequences, a target gene is operably linked to the

control sequences in the proper reading frame. Promoter sequences that can be incorporated into plasmid vectors, and which can support the transcription of the mutant peroxidase gene, include but are not limited to the prokaryotic β -lactamase promoter (Villa-Kamaroff, et al., 1978, Proc. Natl. Acad. Sci. U.S.A. 75:3727-3731) and the tac promoter (DeBoer, et al., 1983, Proc. Natl. Acad. Sci. U.S.A. 80:21-25). Further references can also be found in "Useful proteins from recombinant bacteria" in Scientific American, 1980, 242:74-94.

10 According to one embodiment, B. subtilis is transformed by an expression vector carrying the mutated DNA. If expression is to take place in a secreting microorganism such as B. subtilis, a signal sequence may follow the translation initiation signal and precede the DNA sequence of interest. The
15 signal sequence acts to transport the expression product to the cell wall where it is cleaved from the product upon secretion. The term "control sequences" as defined above is intended to include a signal sequence, when present.

The host organism transformed with the DNA sequence
20 encoding the peroxidase variant of the invention may also be a yeast, preferably a strain of Saccharomyces, e.g. Saccharomyces cerevisiae or Schizosaccharomyces pombe, or Pichia, e.g. Pichia pastoris.

In a currently preferred method of producing the
25 peroxidase variant of the invention, a filamentous fungus is used as the host organism. The filamentous fungus host organism may conveniently be one which has previously been used as a host for producing recombinant proteins, e.g. a strain of Aspergillus sp., such as A. niger, A. nidulans or A. oryzae.
30 The use of A. oryzae in the production of recombinant proteins is extensively described in, e.g. EP 238 023.

For expression of peroxidase variants in Aspergillus, the DNA sequence coding for the peroxidase variant is preceded by a promoter. The promoter may be any DNA sequence exhibiting
35 a strong transcriptional activity in Aspergillus and may be derived from a gene encoding an extracellular or intracellular protein such as an amylase, a glucoamylase, a protease, a

lipase, a peroxidase, a cellulase or a glycolytic enzyme.

Examples of suitable promoters are those derived from the gene encoding A. oryzae TAKA amylase, Rhizomucor miehei aspartic proteinase, A. niger neutral α -amylase, A. niger acid stable α -amylase, A. niger glucoamylase, Rhizomucor miehei lipase, A. oryzae alkaline protease or A. oryzae triose phosphate isomerase.

In particular when the host organism is A. oryzae, a preferred promoter for use in the process of the present invention is the A. oryzae TAKA amylase promoter as it exhibits a strong transcriptional activity in A. oryzae. The sequence of the TAKA amylase promoter appears from EP 238 023.

Termination and polyadenylation sequences may suitably be derived from the same sources as the promoter.

15 The techniques used to transform a fungal host cell may suitably be as described in EP 238 023.

To ensure secretion of the peroxidase variant from the host cell, the DNA sequence encoding the peroxidase variant may be preceded by a signal sequence which may be a naturally occurring signal sequence or a functional part thereof or a synthetic sequence providing secretion of the protein from the cell. In particular, the signal sequence may be derived from a gene encoding an Aspergillus sp. amylase or glucoamylase, a gene encoding a Rhizomucor miehei lipase or protease, or a gene encoding a Humicola cellulase, xylanase or lipase. The signal sequence is preferably derived from the gene encoding A. oryzae TAKA amylase, A. niger neutral α -amylase, A. niger acid-stable α -amylase, Coprinus cinereus or macrorhizus peroxidase, or A. niger glucoamylase.

30 The medium used to culture the transformed host cells may be any conventional medium suitable for growing Aspergillus cells. The transformants are usually stable and may be cultured in the absence of selection pressure. However, if the transformants are found to be unstable, a selection marker introduced into the cells may be used for selection.

The mature peroxidase protein secreted from the host cells may conveniently be recovered from the culture medium by

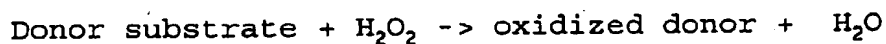
well-known procedures including separating the cells from the medium by centrifugation or filtration, and precipitating proteinaceous components of the medium by means of a salt such as ammonium sulphate, followed by chromatographic procedures such as ion exchange chromatography, affinity chromatography, or the like.

To obtain a bleaching effect of the peroxidase variant, hydrogen peroxide or a precursor of hydrogen peroxide, preferably perborate or percarbonate, or a hydrogen peroxide generating enzyme system, e.g. an oxidase and its substrate, or a peroxycarboxylic acid or a salt thereof, should be present in the bleaching composition of the invention as substrate for the peroxidase variant.

While the mechanism of peroxidase bleaching of coloured substances present on fabrics or in the wash liquor has not yet been elucidated, it is currently believed that the enzyme acts by reducing hydrogen peroxide and oxidizing the coloured substance (electron donor substrate), thereby either generating a colourless substance or providing a substance which is not adsorbed to the fabric. This reaction is shown in Reaction Scheme 1 below

Reaction Scheme 1:

Peroxidase



By using a peroxidase variant according to the invention which is less sensitive to hydrogen peroxide, it may be possible to add a smaller amount of the enzyme to the bleaching/washing liquor and yet obtain a satisfactory bleaching effect.

In the bleaching composition, the amount of peroxidase variant corresponds to a concentration in the wash liquor of between 0.01 and 20 PODU/ml, and the amount of hydrogen peroxide or hydrogen peroxide precursor or hydrogen peroxide generating enzyme system or percarboxylic acid or a salt

thereof corresponds to a hydrogen peroxide concentration of up to 20 mM H_2O_2 .

Determination of peroxidase activity: 1 peroxidase unit (PODU) is the amount of enzyme that catalyzes the conversion of 1 μ mol hydrogen peroxide per minute at the following analytical conditions: 0.88 mM hydrogen peroxide, 1.67 mM 2,2'-azinobis(3-ethylbenzothiazoline-6-sulfonate), 0.1 M phosphate buffer, pH 7.0, incubated at 30°C, photometrically followed at 418 nm.

10 For use of the present peroxidase variant as a bleaching composition, it has been found that the addition of another oxidizable substrate (for the peroxidase variant of the invention) at the beginning or during the washing and/or rinsing process may enhance the bleaching effect of the
15 peroxidase variant employed. This is thought to be ascribable to the formation of radicals or other oxidized states of this substrate which participate in the bleaching or other modification of the coloured substance. Examples of such oxidizable substrates are organic compounds such as phenolic compounds,
20 e.g. p-hydroxybenzenesulfonate. Other examples of phenolic compounds which may be used for the present purpose are those given in M. Kato and S. Shimizu, Plant Cell Physiol. 26(7), 1985, pp. 1291-1301 (cf. Table 1 in particular) or B.C. Saunders et al., Peroxidase, London, 1964, p. 141 ff. In WO
25 94/12621 other types of enhancing agents are disclosed which may be used for the present purpose, e.g. phenothiazines and phenoxazines and derivatives thereof such as 10-methylphenothiazine, 10-phenothiazine-propionic acid, N-hydroxysuccinimide-
30 10-phenothiazine-propionate, 10-ethyl-4-phenothiazine-carboxylic acid, 10-ethylphenothiazine, 10-propylphenothiazine, 10-isopropylphenothiazine, methyl-10-phenothiazinepropionate, 10-phenylphenothiazine, 10-allylphenothiazine, 10-(3-(4-methyl-1-piperazinyl)propyl)phenothiazine, 10-(2-pyrrolidinoethyl)-
phenothiazine, promazine, 2-chloro-10-methylphenothiazine, 2-acetyl-10-methylphenothiazine and 10-methylphenoxazine.

The amount of oxidizable substrate corresponds to a concentration in the wash liquor of between 0.1 μ M and 100 μ M.

Detergent Compositions

According to the invention, the peroxidase variant may be added as a component of a detergent composition. As such, it may be included in the detergent composition in the form of a detergent additive. The detergent composition as well as the detergent additive may additionally comprise one or more other enzymes, such as proteases, lipases, amylases, cutinases and cellulases.

In a specific aspect, the invention provides a detergent additive. The enzymes may be included in a detergent composition by adding separate additives containing one or more enzymes, or by adding a combined additive comprising all of these enzymes. A detergent additive of the invention, i.e. a separated additive or a combined additive, can be formulated e.g. as granulates, liquids, slurries, etc. Preferred detergent additive formulations are granulates, in particular non-dusting granulates, liquids, in particular stabilized liquids, slurries, or protected enzymes.

Non-dusting granulates may be produced, e.g., as disclosed in US 4,106,991 and 4,661,452 (both to Novo Industri A/S) and may optionally be coated by methods known in the art. Examples of waxy coating materials are poly(ethylene oxide) products (polyethyleneglycol, PEG) with mean molar weights of 1000 to 20000; ethoxylated nonylphenols having from 16 to 50 ethylene oxide units; ethoxylated fatty alcohols in which the alcohol contains from 12 to 20 carbon atoms and in which there are 15 to 80 ethylene oxide units; fatty alcohols; fatty acids; and mono- and di- and triglycerides of fatty acids. Examples of film-forming coating materials suitable for application by fluid bed techniques are given in patent GB 1483591. Liquid enzyme preparations may, for instance, be stabilized by adding a polyol such as propylene glycol, a sugar or sugar alcohol, lactic acid or boric acid according to established methods. Other enzyme stabilizers are well known in the art. Protected enzymes may be prepared according to the method disclosed in EP 238,216.

The detergent composition of the invention may be in

any convenient form, e.g. as powder, granules, paste or liquid. A liquid detergent may be aqueous, typically containing up to 70% of water and 0-30% of organic solvent, or nonaqueous.

The detergent composition comprises one or more surfactants, each of which may be anionic, nonionic, cationic, or zwitterionic. The detergent will usually contain 0-50% of anionic surfactant such as linear alkylbenzenesulfonate (LAS), alpha-olefinsulfonate (AOS), alkyl sulfate (fatty alcohol sulfate) (AS), alcohol ethoxysulfate (AEOS or AES), secondary alkanesulfonates (SAS), alpha-sulfo fatty acid methyl esters, alkyl- or alkenylsuccinic acid or soap. It may also contain 0-40% of nonionic surfactant such as alcohol ethoxylate (AEO or AE), carboxylated alcohol ethoxylates, nonylphenol ethoxylate, alkylpolyglycoside, alkyl dimethylamineoxide, ethoxylated fatty acid monoethanolamide, fatty acid monoethanolamide, or polyhydroxy alkyl fatty acid amide (e.g. as described in WO 92/06154).

The detergent composition may additionally comprise one or more other enzymes, such as amylases, lipases, cutinases, proteases and cellulases.

The detergent may contain 1-65% of a detergent builder or complexing agent such as zeolite, diphosphate, triphosphate, phosphonate, citrate, nitrilotriacetic acid (NTA), ethylenediaminetetraacetic acid (EDTA), diethylenetriaminepentaacetic acid (DTMPA), alkyl- or alkenylsuccinic acid, soluble silicates or layered silicates (e.g. SKS-6 from Hoechst). The detergent may also be unbuilt, i.e. essentially free of detergent builder.

The detergent may comprise one or more polymers. Examples are carboxymethylcellulose (CMC), poly(vinylpyrrolidone) (PVP), polyethyleneglycol (PEG), poly(vinyl alcohol) (PVA), polycarboxylates such as polyacrylates, maleic/acrylic acid copolymers and lauryl methacrylate/acrylic acid copolymers.

The detergent may contain a bleaching system which may comprise a H_2O_2 source such as perborate or percarbonate which may be combined with a peracid-forming bleach activator

such as tetraacetythylenediamine (TAED) or nonanoyloxybenzenesulfonate (NOBS). Alternatively, the bleaching system may comprise peroxyacids of e.g. the amide, imide, or sulfone type.

The enzymes of the detergent composition of the invention may be stabilized using conventional stabilizing agents, e.g. a polyol such as propylene glycol or glycerol, a sugar or sugar alcohol, lactic acid, boric acid, or a boric acid derivative as e.g. an aromatic borate ester, and the composition may be formulated as described in e.g. WO 92/19709 and 10WO 92/19708.

The detergent may also contain other conventional detergent ingredients such as e.g. fabric conditioners including clays, foam boosters, suds suppressors, anti-corrosion agents, soil-suspending agents, anti-soil redeposition agents, 15dyes, bactericides, optical brighteners, or perfume.

The pH (measured in aqueous solution at use concentration) will usually be neutral or alkaline, e.g. 7-11.

Particular forms of detergent compositions within the scope of the invention include:

201) A detergent composition formulated as a granulate having a bulk density of at least 600 g/l comprising

- linear alkylbenzenesulfonate (calculated as acid)	7 - 12%
25 - alcohol ethoxysulfate (e.g. C ₁₂₋₁₈ alcohol, 1-2 EO) or alkyl sulfate (e.g. C ₁₆₋₁₈)	1 - 4%
- alcohol ethoxylate (e.g. C ₁₄₋₁₅ alcohol, 7 EO)	5 - 9%
- sodium carbonate (as Na ₂ CO ₃)	14 - 20%
30- soluble silicate (as Na ₂ O, 2SiO ₂)	2 - 6%
- zeolite (as NaAlSiO ₄)	15 - 22%
- sodium sulfate (as Na ₂ SO ₄)	0 - 6%
- sodium citrate/citric acid (as C ₆ H ₅ Na ₃ O ₇ /C ₆ H ₈ O ₇)	0 - 15%
35- sodium perborate (as NaBO ₃ ·H ₂ O)	11 - 18%
- TAED	2 - 6%

- carboxymethylcellulose 0 - 2%
- polymers (e.g. maleic/acrylic acid copolymer, PVP, PEG) 0 - 3%
- enzymes 0 - 5%
- 5- minor ingredients (e.g. suds suppressors, perfume, optical brightener, photobleach) 0 - 5%

2) A detergent composition formulated as a granulate having a bulk density of at least 600 g/l comprising

- 10- linear alkylbenzenesulfonate (calculated as acid) 6 - 11%
- alcohol ethoxysulfate (e.g. C₁₂₋₁₈ alcohol, 1-2 EO) or alkyl sulfate (e.g. C₁₆₋₁₈) 1 - 3%
- 15- alcohol ethoxylate (e.g. C₁₄₋₁₅ alcohol, 7 EO) 5 - 9%
- sodium carbonate (as Na₂CO₃) 15 - 21%
- soluble silicate (as Na₂O, 2SiO₂) 1 - 4%
- zeolite (as NaAlSiO₄) 24 - 34%
- 20- sodium sulfate (as Na₂SO₄) 4 - 10%
- sodium citrate/citric acid (as C₆H₅Na₃O₇/C₆H₈O₇) 0 - 15 %
- carboxymethylcellulose 0 - 2%
- polymers (e.g. maleic/acrylic acid copolymer, PVP, PEG) 1 - 6%
- 25 - enzymes 0 - 5%
- minor ingredients (e.g. suds suppressors, perfume) 0 - 5%

3) A detergent composition formulated as a granulate having a bulk density of at least 600 g/l comprising

- linear alkylbenzenesulfonate (calculated as acid) 5 - 9%
- alcohol ethoxylate (e.g. C₁₂₋₁₅ alcohol, 7 EO) 7 - 14%

- soap as fatty acid (e.g. C ₁₆₋₂₂)	1 - 3%
- sodium carbonate (as Na ₂ CO ₃)	10 - 17%
- soluble silicate (as Na ₂ O, 2SiO ₂)	3 - 9%
5- zeolite (as NaAlSiO ₄)	23 - 33%
- sodium sulfate (as Na ₂ SO ₄)	0 - 4%
- sodium perborate (as NaBO ₃ ·H ₂ O)	8 - 16%
- TAED	2 - 8%
- phosphonate (e.g. EDTMPA)	0 - 1%
10- carboxymethylcellulose	0 - 2%
- polymers (e.g. maleic/acrylic acid copolymer, PVP, PEG)	0 - 3%
- enzymes	0 - 5%
- minor ingredients (e.g. suds suppressors, 15 perfume, optical brightener)	0 - 5%

4) A detergent composition formulated as a granulate having a bulk density of at least 600 g/l comprising

- linear alkylbenzenesulfonate (calculated as acid)	8 - 12%
20- alcohol ethoxylate (e.g. C ₁₂₋₁₅ alcohol, 7 EO)	10 - 25%
- sodium carbonate (as Na ₂ CO ₃)	14 - 22%
- soluble silicate (as Na ₂ O, 2SiO ₂)	1 - 5%
- zeolite (as NaAlSiO ₄)	25 - 35%
25- sodium sulfate (as Na ₂ SO ₄)	0 - 10%
- carboxymethylcellulose	0 - 2%
- polymers (e.g. maleic/acrylic acid copolymer, PVP, PEG)	1 - 3%
- enzymes	0 - 5%
30- minor ingredients (e.g. suds suppressors, perfume)	0 - 5%

- 5) An aqueous liquid detergent composition comprising
- linear alkylbenzenesulfonate
(calculated as acid) 15 - 21%
 - alcohol ethoxylate
 - 5 (e.g. C₁₂₋₁₅ alcohol, 7 EO or C₁₂₋₁₅ alcohol, 5 EO) 12 - 18%
 - soap as fatty acid (e.g. oleic acid) 3 - 13%
 - alkenylsuccinic acid (C₁₂₋₁₄) 0 - 13%
 - aminoethanol 8 - 18%
 - 10- citric acid 2 - 8%
 - phosphonate 0 - 3%
 - polymers (e.g. PVP, PEG) 0 - 3%
 - borate (as B₄O₇) 0 - 2%
 - ethanol 0 - 3%
 - 15- propylene glycol 8 - 14%
 - enzymes 0 - 5%
 - minor ingredients
(e.g. dispersants, suds suppressors,
perfume, optical brightener) 0 - 5%
- 206) An aqueous structured liquid detergent composition comprising
- linear alkylbenzenesulfonate
(calculated as acid) 15 - 21%
 - alcohol ethoxylate
 - 25 (e.g. C₁₂₋₁₅ alcohol, 7 EO or C₁₂₋₁₅ alcohol, 5 EO) 3 - 9%
 - soap as fatty acid (e.g. oleic acid) 3 - 10%
 - zeolite (as NaAlSiO₄) 14 - 22%
 - potassium citrate 9 - 18%
 - 30- borate (as B₄O₇) 0 - 2%
 - carboxymethylcellulose 0 - 2%
 - polymers (e.g. PEG, PVP) 0 - 3%

- anchoring polymers as
e.g. lauryl methacrylate/acrylic acid copolymer;
molar ratio 25:1; MW 3800 0 - 3%
 - glycerol 0 - 5%
 - 5- enzymes 0 - 5%
 - minor ingredients
(e.g. dispersants, suds suppressors, perfume,
optical brighteners) 0 - 5%
- 7) A detergent composition formulated as a granulate having a
bulk density of at least 600 g/l comprising
- fatty alcohol sulfate 5 - 10%
 - ethoxylated fatty acid monoethanolamide 3 - 9%
 - soap as fatty acid 0 - 3%
 - sodium carbonate (as Na_2CO_3) 5 - 10%
 - 15- soluble silicate (as $\text{Na}_2\text{O}, 2\text{SiO}_2$) 1 - 4%
 - zeolite (as NaAlSiO_4) 20 - 40%
 - sodium sulfate (as Na_2SO_4) 2 - 8%
 - sodium perborate (as $\text{NaBO}_3 \cdot \text{H}_2\text{O}$) 12 - 18%
 - TAED 2 - 7%
 - 20- polymers (e.g. maleic/acrylic acid copolymer,
PEG) 1 - 5%
 - enzymes 0 - 5%
 - minor ingredients (e.g. optical brightener,
suds suppressors, perfume) 0 - 5%
- 258) A detergent composition formulated as a granulate comprising
- linear alkylbenzenesulfonate
(calculated as acid) 8 - 14%
 - ethoxylated fatty acid monoethanolamide 5 - 11%
 - soap as fatty acid 0 - 3%
 - 30- sodium carbonate (as Na_2CO_3) 4 - 10%
 - soluble silicate (as $\text{Na}_2\text{O}, 2\text{SiO}_2$) 1 - 4%
 - zeolite (as NaAlSiO_4) 30 - 50%

- sodium sulfate (as Na_2SO_4)	3 - 11%
- sodium citrate (as $\text{C}_6\text{H}_5\text{Na}_3\text{O}_7$)	5 - 12%
- polymers (e.g. PVP, maleic/acrylic acid copolymer, PEG)	1 - 5%
5- enzymes	0 - 5%
- minor ingredients (e.g. suds suppressors, perfume)	0 - 5%

9) A detergent composition formulated as a granulate comprising

10 - linear alkylbenzenesulfonate (calculated as acid)	6 - 12%
- nonionic surfactant,	1 - 4%
- soap as fatty acid	2 - 6%
- sodium carbonate (as Na_2CO_3)	14 - 22%
- zeolite (as NaAlSiO_4)	18 - 32%
15- sodium sulfate (as Na_2SO_4)	5 - 20%
- sodium citrate (as $\text{C}_6\text{H}_5\text{Na}_3\text{O}_7$)	3 - 8%
- sodium perborate (as $\text{NaBO}_3 \cdot \text{H}_2\text{O}$)	4 - 9%
- bleach activator (e.g. NOBS or TAED)	1 - 5%
- carboxymethylcellulose	0 - 2%
20- polymers (e.g. polycarboxylate or PEG)	1 - 5%
- enzymes	0 - 5%
- minor ingredients (e.g. optical brightener, perfume)	0 - 5%

10) An aqueous liquid detergent composition comprising

25- linear alkylbenzenesulfonate (calculated as acid)	15 - 23%
- alcohol ethoxysulfate (e.g. C_{12-15} alcohol, 2-3 EO)	8 - 15%
- alcohol ethoxylate (e.g. C_{12-15} alcohol, 7 EO or C_{12-15} alcohol, 5 EO)	3 - 9%

- soap as fatty acid (e.g. lauric acid)	0 - 3%
- aminoethanol	1 - 5%
- sodium citrate	5 - 10%
- hydrotrope (e.g. sodium toluenesulfonate)	2 - 6%
5- borate (as B_4O_7)	0 - 2%
- carboxymethylcellulose	0 - 1%
- ethanol	1 - 3%
- propylene glycol	2 - 5%
- enzymes	0 - 5%
10- minor ingredients (e.g. polymers, dispersants, perfume, optical brighteners)	0 - 5%

11) An aqueous liquid detergent composition comprising

- linear alkylbenzenesulfonate (calculated as acid)	20 - 32%
15- alcohol ethoxylate (e.g. C_{12-15} alcohol, 7 EO or C_{12-15} alcohol, 5 EO)	6 - 12%
- aminoethanol	2 - 6%
- citric acid	8 - 14%
20- borate (as B_4O_7)	1 - 3%
- polymer (e.g. maleic/acrylic acid copolymer, anchoring polymers as e.g. lauryl methacrylate/acrylic acid copolymer and CMC)	0 - 3%
25- glycerol	3 - 8%
- enzymes	0 - 5%
- minor ingredients (e.g. hydrotropes, dispersants, perfume, optical brighteners)	0 - 5%

12) A detergent composition formulated as a granulate having a
30 bulk density of at least 600 g/l comprising

- anionic surfactant (linear alkylbenzenesulfonate, alkyl sulfate, alpha-

olefinsulfonate, alpha-sulfo fatty acid methyl esters, alkanesulfonates, soap)	25 - 40%
- nonionic surfactant (e.g. alcohol ethoxylate)	1 - 10%
5- sodium carbonate (as Na_2CO_3)	8 - 25%
- soluble silicates (as Na_2O , 2SiO_2)	5 - 15%
- sodium sulfate (as Na_2SO_4)	0 - 5%
- zeolite (as NaAlSiO_4)	15 - 28%
- sodium perborate (as $\text{NaBO}_3 \cdot 4\text{H}_2\text{O}$)	0 - 20%
10- bleach activator (TAED or NOBS)	0 - 5%
- enzymes	0 - 5%
- minor ingredients (e.g. perfume, optical brighteners)	0 - 3%

13) Detergent formulations as described in 1) - 12) where the content of linear alkylbenzenesulfonate - or a part of it - is substituted by alkyl sulfate (C_{12} - C_{18}).

14) Detergent formulations as described in 1) - 13) which contain a stabilized or encapsulated peracid either as an additional component or as a substitute for already specified bleach systems.

15) Detergent compositions as described in 1), 3), 7), 9) and 12) where the content of perborate is substituted with percarbonate.

16) Detergent compositions as described in 1), 3), 7), 9) and 12) which additionally contains a Manganese catalyst. The Manganese catalyst may e.g. be one of the compounds described in "Efficient manganese catalysts for low-temperature bleaching", Nature 369, 1994, pp. 637-639.

17) Detergent composition formulated as a nonaqueous detergent solid comprising a liquid nonionic surfactant as e.g. linear

alkoxylated primary alcohol, a builder system (e.g. phosphate), enzyme and alkali. The detergent may also comprise anionic surfactant and/or a bleach system.

It is at present contemplated that, in the detergent composition of the invention, the peroxidase variant may be added in an amount corresponding to a concentration in the wash liquor of between 0.01 to 20 PODU/ml.

The present invention is further illustrated in the following examples which are not in any way intended to limit the scope of the invention as claimed.

EXAMPLE 1

Construction of a plasmid expressing the K48S variant of *Coprinus cinereus* peroxidase

1. Cloning of cDNA encoding a *Coprinus cinereus* peroxidase

15 Construction of a Probe by PCR

Peroxidase cDNA fragments were prepared by polymerase chain reaction (PCR) using specific oligonucleotide primers (R.K. Saiki et al., Science 239, 1988, pp. 487-491) constructed on the basis of the amino acid sequence of the *Coprinus* 20 macrorrhizus peroxidase. PCR was carried out using the Gene Amp kit and apparatus (available from Perkin Elmer Cetus, Norwalk, CT, USA) in accordance with the manufacturer's instructions, with the exception that the reaction was conducted at 28°C for the first three cycles in order to obtain better hybridisation 25 to the first strand cDNA (prepared from mRNA obtained from *Coprinus cinereus*, IFO 8371) and subsequently at 65°C for 30 cycles of PCR.

The following specific primers were used for PCR:

1. 5'-GCGCGAATTTCGTNGGNATNAACCACGG-3' ^{T T}
2. 3'-TACAGNTTGACGGGNGGCCTAGGCG-5' ^{A A}
- 5 3. 5'-GCGAATTCACNCCNCAGGTNTTCGACAC-3' ^{A T T}
4. 3'-GGNAAGGGNCCNCTCAAGCCTAGGCG-5' ^{A T A}
- 10 5. 5'-GCGCGAATTCTGGCAGTCNAC-3' ^A
6. 5'-GCGCGAATTCTGGCAGAGNATG-3' ^A
7. 3'-CGNTACCGNTTCTACAGCCTAGG-5' ^T

¹⁵"N" denoting a mixture of all four nucleotides.

The primers were combined as follows: 1 with 2, 3 with 4, 5 with 7, 6 with 7, 1 with 4, 1 with 7 and 3 with 7. The PCR fragments were thus extended with an EcoRI site at the 5'-end and a BamHI site at the 3'-end. The PCR reactions were analyzed on a 1% agarose gel. Bands of the expected size were found in all reactions. To verify that the bands corresponded to peroxidase-specific sequences, the gel was subjected to Southern blotting and hybridised to an oligonucleotide probe with the following sequence

25 ^{T A A A T}
5'-GTCTCGATGTAGAACTG-3'
^T

which is positioned between PCR primers 3 and 4. The probe was found to hybridise to bands of approximately 130 bp, 420 bp, 30540 bp and 240 bp, thus confirming that the DNA bands observed correspond to peroxidase sequences.

DNA from the various PCR reactions was digested with EcoRI and BamHI and cloned into the plasmid pUC19 (C. Yanisch-

Perron et al., Gene **33**, 1985, pp. 103-119). Colonies containing the correct PCR fragments were identified by hybridisation using the oligonucleotide probe specified above. DNA from positive colonies was analyzed by restriction enzyme mapping and partial DNA sequence analysis as described by Sanger et al., Proc. Natl. Acad. Sci. USA **74**, 1977, pp. 5463-5467. A 430 bp fragment from one of the clones, obtained by using primer 1 and 4, was used to screen a Coprinus cinereus cDNA library as described below.

10 Construction of a Coprinus cinereus cDNA library in E. coli

Total RNA was extracted from homogenized Coprinus cinereus (IFO 8371) mycelium, collected at the time for maximum activity of the peroxidase by methods as described by Boel et al. (EMBO J., **3**: 1097-1102, 1984) and Chirgwin et al. (Biochemistry (Wash), **18**: 5294-5299, 1979). Poly(A)-containing RNA is obtained by two cycles of affinity chromatography on oligo(dT)-cellulose as described by Aviv and Leder (PNAS, USA **69**:1408-1412, 1972). cDNA is synthesized by means of a cDNA synthesis kit from Invitrogen according to the manufacturer's instructions. About 50.000 E. coli recombinants from the Coprinus cinereus cDNA library were transferred to Whatman 540 paper filters. The colonies were lysed and immobilized as described by Gergen et al. (Nucleic Acids Res. **7**, 2115-2135, 1979). The filters were hybridized with the ³²P-labelled 430 bp peroxidase-specific probe in 0.2 x SSC, 0.1% SDS. Hybridization and washing of the filters were conducted at 65°C followed by autoradiography for 24 hours with an intensifier screen. After autoradiography, the filters were washed at increasing temperatures followed by autoradiography for 24 hours with an intensifier screen. In this way, more than 50 positive clones were identified. Miniprep plasmid DNA was isolated from hybridizing colonies by standard procedures (Birnboim and Doly Nucleic Acids Res. **7**, 1513-1523, 1979), and the DNA sequence of the cDNA insert was determined by the Sanger dideoxy procedure (Sanger et al., Proc. Natl. Acad. Sci. USA **74**, 1977, pp. 5463-5467). The peroxidase cDNA fragment was excised from the vector

by cleavage with HindIII/XhoI and was purified by agarose gel electrophoresis, electroeluted and made ready for ligation reactions. The cDNA fragment was ligated to HindIII/XhoI digested pHD414 to generate pCiP in which the cDNA is under transcriptional control of the TAKA promotor from Aspergillus oryzae and the AMG terminator from Aspergillus niger.

Construction of the Aspergillus expression vector pHD414

The vector pHD414 is a derivative of the plasmid p775 (described in EP 238 023). In contrast to p775, pHD414 has a string of unique restriction sites between the promotor and the terminator.

The plasmid was constructed by removal of an approximately 200 bp long fragment (containing undesirable restriction sites) at the 3' end of the terminator, and subsequent removal of an approximately 250 bp long fragment at the 5' end of the promotor, also containing undesirable restriction sites. The 200 bp region was removed from p775 by cleavage with NarI (positioned in the pUC vector) and XbaI (positioned just 3' to the terminator), subsequent filling in the generated ends with Klenow DNA polymerase + dNTP, purification of the vector fragment on gel and religation of the vector fragment. The DNA was transformed into E. coli MC1061 as described above. 10 colonies (pHD413-1 to -10) were selected and analyzed by restriction enzyme analysis. One of the clones exhibiting the expected band pattern in the restriction enzyme analysis was used in the construction of pHD414.

pHD413 was cut with StuI (positioned in the 5' end of the promoter) and PvuII (positioned in the pUC vector) and fractionated on a gel. The vector fragment was purified, religated and transformed into E. coli MC1061. 12 colonies were selected and analyzed by restriction enzyme analysis. All 12 clones exhibited the expected band pattern. The plasmid pHD414 is shown in Fig. 1.

2. 3-step PCR mutagenesis:

3-step mutagenisation involves the use of four primers:

Mutagenisation primer (=A): 5'-CCT GTT CGA TCG ATT CTT AGA-3'

PCR Helper 1 (=B): 5'-TGA TCA TAG TAC CAT CTA ATT ACA TCA AGC
5 GGC-3'

PCR Helper 2 (=C): 5'-CTG TAA TAC GAC TCA CTA-3'

PCR Handle (=D): 5'-TGA TCA GAC TAG TAC CAT-3'

Primer A and B were diluted to 20 pmole/ μ l. Primer C and D were diluted to 100 pmole/ μ l.

10 All 3 steps were carried out in a 10 x PCR buffer containing: 100 mM Tris-HCl, pH 8.3, 500 mM KCl, 15 mM MgCl₂, 0.1% gelatin, 200 μ l of each of 2 mM dATP, 2 mM dCTP, 2 mM dGTP, 2 mM TTP, and 200 μ l of H₂O.

In step 1, a reaction mixture composed of 10 μ l of 10x PCR buffer, 50 μ l of 2 x nucleotide solution, 5 μ l of primer A, 5 μ l of primer B, 1 μ l of pCiP (0.05 μ g/ μ l), 30 μ l of H₂O, 0.5 μ l of Taq polymerase, and 80 μ l of paraffin, was run through 1 cycle consisting of 2 minutes at 94°C, 15 cycles consisting of 1 minute at 94°C, 1 minute at 50°C and 2 minutes at 72°C, 15 cycles consisting of 1 minute at 94°C, 1 minute at 50°C and 3 minutes at 72°C, and 1 cycle consisting of 5 minutes at 72°C.

10 μ l of the PCR product were purified on an agarose gel and redissolved in 10 μ l of H₂O. Then, step 2 was carried out. A reaction mixture composed of 10 μ l of 10 x PCR buffer, 50 μ l of 2 x nucleotide solution, 5 μ l of the purified product of step 1, 1 μ l pCiP (0.05 μ g/ μ l), 30 μ l of H₂O, 0.5 μ l of Taq polymerase, and 80 μ l of paraffin, was run through 1 cycle consisting of 5 minutes at 94°C, 2 minutes at 50°C and 10 minutes at 72°C.

To the step 2 reaction mixture, 1 μ l of primer C and

1 μ l of primer D were added. The PCR reaction was carried out as described for step 1.

3. Isolation of the mutated restriction fragment:

The product from step 3 was isolated from an agarose gel and re-dissolved in 20 μ l H₂O. Then, it was digested with the restriction enzymes XbaI and KpnI in NEBuffer 2 (New England Biolabs) supplemented with bovine serum albumin (BSA) in a total volume of 20 μ l at 37°C overnight. The 570 bp XbaI/KpnI fragment was isolated from an agarose gel.

104. Ligation to expression vector pCiP:

The expression plasmid pCiP was cleaved with XbaI and KpnI under the conditions indicated above, and the large fragment was isolated from an agarose gel. To this vector, the mutated fragment isolated above was ligated and the ligation mixture was used to transform E.coli. The presence and orientation of the fragment were verified by cleavage of a plasmid preparation from a transformant with restriction enzymes. Sequence analysis was carried out on the double-stranded plasmid using the di-deoxy chain termination procedure developed by Sanger. The resulting plasmid is identical to pCiP except for the altered codon.

EXAMPLE 2

Construction of plasmids expressing other variants of Coprinus peroxidase

25 The following mutants were constructed using the same method as described in Example 1, except that other restriction enzymes were used for digesting the PCR-product and the vector used for recloning the mutated fragment. Mutations and primers used for the modifications are listed below.

Mutation	Primer A sequence
K48S	5'-CCT GTT CGA TCG ATT CTT AGA-3'
V53K	5'-CTT AGA ATT AAA TTC CAT GAC-3'
G72Q	5'-GAT GGA GCC ATC GGC GCC TCC TTG ACC GAA TTG ACC-3'
5A91C	5'-GCC TTC CCG TGC AAT GGC GGC-3'
N92K	5'-TTC CCG GCT AAA GGA GGC CTC-3'
H109K	5'-GGT ATT AAT AAA GGT GTC TCT-3'
Q118E	5'-GAT CTC ATC GAA TTC GCC ACT-3'
M125G	5'-GCC GTC GGC GGG TCC AAC TGC-3'
10M125A	5'-GCC GTC GGC GCC TCC AAC TGC-3'
S147Q	5'-ACC GGG GAT CAA GCT TGG AGG TTG GGG TTG GGA ACT-3'
I152C	5'-CCT TCG TTG TGT CCC GGG CCC-3'
P155C	5'-G ATC CCC GGG TGC GGA AAC ACT-3'
M166G	5'-TTG GAT CGT GGC GGC GAT GCA-3'
15N192K	5'-GAG GGT TTA AAA TCG GCC ATC-3'
I195K	5'-G AAC TCG GCC AAA TTC AGG TCT-3'
V206R	5'-CTG GGT ATC GAA GCG CTG AGG GGT CGA-3'
K218R	5'-CTG AGT GGT GCC TCG GAG CAA GGT CTC-3'
E214L	5'-TCT ACA TTT TAA CCT TGC TC-3'
20F229G	5'-GAG CTC CTC GGC GCC GCC GAG AGA AGG-3'
A230C	5'-CTC GGC TTT TGC GAG GAA CTC-3'
E239G	5'-TTC CCT GGC GGC TTC CGC ATG-3'
E239H	5'-TTC CCT GGC CAC TTC CGC ATG-3'
E239K	5'-TTC CCT GGC AAA TTC CGC ATG-3'
25E239L	5'-TTC CCT GGC CTA TTC CGC ATG-3'
E239M	5'-TTC CCT GGC ATG TTC CGC ATG-3'
E239Q	5'-TTC CCT GGC CAA TTC CGC ATG-3'
E239S	5'-TTC CCT GGC TCA TTC CGC ATG-3'
E239T	5'-TTC CCT GGC ACA TTC CGC ATG-3'
30E239W	5'-TTC CCT GGC TGG TTC CGC ATG-3'
E239R	5'-TTC CCT GGC CGA TTC CGC ATG-3'
M242G	5'-GAA TCC CGC GGG AGG TCC GAT-3'
S244C	5'-T CGC ATG AGG TGC GAT GCT CTC-3'
S252P	5'-CG GCA GGC GGT CCG CGG GTC GCG AGC-3'
35W258F	5'-GCA TGC CGA TTT CAA TCC AT-3'
W258H	5'-GCA TGC CGA CAT CAA TCC AT-3'

M261F	5'-TGG CAA TCC TTT ACT AGT AGC-3'
M261G	5'-TGG CAA TCC GGG ACC AGC AGC-3'
M261Y	5'-TGG CAA TCC TAT ACC AGC AGC-3'
M261V	5'-TGG CAA TCC GTC ACC AGC AGC-3'
5M268L	5'-AAT GAA GTC CTA GGC CAG CGA-3'
M268G	5'-AAT GAA GTT GGG GGC CAG CGA-3'
M268A	5'-AAT GAA GTT GCA GGC CAG CGA-3'
M268F	5'-AAT GAA GTT TTT GGC CAG CGA-3'
Y272F	5'-GGG CCA GCG CTT TCG CGC CGC C-3'
10Y272G	5'-GGC CAG CGA GGG CGC GCC GCC-3'
Y272A	5'-GGC CAG CGA GCC CGC GCC GCC-3'
M276S	5'-CGC GCC GCC TTT GCC AAG ATG-3'
M276I	5'-CGC GCC GCC ATA GCC AAG ATG-3'
M276H	5'-CGC GCC GCC CAC GCC AAG ATG-3'
15K278R	5'-AG AAC AGA CAT GCG CGC CAT GGC GGC-3'
M279G	5'-ATG GCC AAG GGG TCT GTT CTC-3'
M279A	5'-ATG GCC AAG GCC TCT GTT CTC-3'
A304E	5'-AT AAC AGG CGC CTC GTT GGA CAC-3'
V314P	5'-GGC CTT ACT CCC GAT GAT ATC-3'
20K41R + K48R	5'-AAT TCT AAG AAT TCG GCG AAC AGG GCT CTC ACA TCG GGA CCC TTG-3'
G167N + V176L	5'-AAG CAA GTC AAC GAG CTC ATC AGG GCT GAA GCC TGC ATC GTT CAT ACG ATC CAA-3'
R241E + E239K	5'-TTC CCT GGC AAG TTC GAA ATG AGG TCC-3'.

25 It should be noted that variants in position 1-29 were digested with BamHI and XbaI in NEBuffer 3 (New England Biolabs) supplemented with BSA, resulting in a 160 bp fragment. Variants in position 30-219 were digested with XbaI/KpnI in NEBuffer 2 supplemented with BSA, resulting in a 570 bp fragment. Variants in position 220-277 were digested with KpnI/MscI in NEBuffer 2 supplemented with BSA, resulting in a 170 bp fragment. Variants in position 278-363 were digested with MscI/XhoI in NEBuffer 2 supplemented with BSA, resulting in a 420 bp fragment.

EXAMPLE 3

Expression of Coprinus peroxidase variants in Aspergillus oryzaeTransformation of Aspergillus oryzae or Aspergillus niger (general procedure)

100 ml of YPD medium (Sherman et al., Methods in Yeast Genetics, Cold Spring Harbor Laboratory, 1981) were inoculated with spores of A. oryzae or A. niger and incubated with shaking at 37°C overnight. The mycelium was harvested by filtration through miracloth and washed with 200 ml of 0.6 M MgSO₄. The mycelium was suspended in 15 ml of 1.2 M MgSO₄, 10 mM NaH₂PO₄, pH = 5.8. The suspension was cooled on ice, and 1 ml of buffer containing 120 mg of Novozym[®] 234, batch 1687 was added. After 5 minutes 1 ml of 12 mg/ml BSA (Sigma type H25) was added, and incubation with gentle agitation was continued for 1.5-2.5 hours at 37°C until a large number of protoplasts was visible in a sample inspected under the microscope.

The suspension was filtered through miracloth, the filtrate was transferred to a sterile tube and overlaid with 205 ml of 0.6 M sorbitol, 100 mM Tris-HCl, pH = 7.0. Centrifugation was performed for 15 minutes at 100 x g, and protoplasts were collected from the top of the MgSO₄ cushion. 2 volumes of STC (1.2 M sorbitol, 10 mM Tris-HCl, pH = 7.5, 10 mM CaCl₂) were added to the protoplast suspension and the mixture was centrifuged for 5 minutes at 1000 x g. The protoplast pellet was resuspended in 3 ml of STC and repelleted. This procedure was repeated. Finally the protoplasts were resuspended in 0.2-1 ml of STC.

100 µl of the protoplast suspension were mixed with 305-25 µg of the appropriate DNA in 10 µl of STC. Protoplasts from the A1560 strain of A. oryzae (IFO 4177) were mixed with pToC186 (an A. nidulans amdS gene carrying plasmid). The mixture was left at room temperature for 25 minutes. 0.2 ml of 60% PEG 4000 (BDH 29576), 10 mM CaCl₂ and 10 mM Tris-HCl, pH =

7.5, were added and carefully mixed (twice) and finally 0.85 ml of the same solution was added and carefully mixed. The mixture was left at room temperature for 25 minutes, spun at 2500 x g for 15 minutes and the pellet was resuspended in 2 ml of 1.2 M sorbitol. After another sedimentation, the protoplasts were spread on the appropriate plates. Protoplasts from the A1560 strain transformed with pToC186 were spread on minimal plates (Cove Biochem. Biophys. Acta 113 (1966) 51-56) containing 1.0 M sucrose, pH = 7.0, 10 mM acetamide as nitrogen source and 20 mM CsCl to inhibit background growth. After incubation for 4-7 days at 37°C spores were picked, suspended in sterile water and spread for single colonies. This procedure was repeated and spores of a single colony after the second reisolation were stored as defined transformants.

15Production of recombinant Coprinus cinereus peroxidase variants in an A. oryzae strain

Plasmids containing the appropriate mutations in the peroxidase gene were transformed into A. oryzae A1560 by cotransformation with pToC186 containing the amdS gene from A. nidulans as described above with a mixture of equal amounts of pCiP and pToC186 (approximately 5 µg of each). Transformants which were able to use acetamide as their sole nitrogen source were reisolated twice. After growth on YPD medium (Sherman et al. 1981) for three days culture supernatants were analyzed by 25a peroxidase activity assay (PODU).

EXAMPLE 4

Hydrogen Peroxide Stability

Wild-type recombinant Coprinus cinereus peroxidase (rCiP) and the variants E239K, Y272F, M276S, M242I + Y272F + 30E239K prepared as described above, were tested for their stability in the presence of H₂O₂.

Samples of the variants tested were purified in the

following way:

5 l of culture broth were clarified by suction-filtration through Propex cloth. 3.5 l were added 1378 g of ammonium sulphate (final concentration 2.5 M) with gentle stirring. The mixture was left at 5°C for 16 h. The precipitate was collected by centrifugation at 2500*g for 30 min at 5°C. The precipitate was redissolved in a CaCl_2 solution (5 mM), final volume was 200 ml. 30 ml of this solution were dialysed against a 12.5 mM Bis-tris pH 6.0 buffer; 5 l for 4 h and 5 l for 16 h. Final volume was 68 ml.

23 ml of dialysate were applied to a Q-Sepharose HP column (Pharmacia, 26 mm Ø, 100 mm bed height). The column was equilibrated with 12.5 mM Bis-tris pH 6.0 buffer. Flow rate 4 ml/min. 400 ml of starting buffer were passed over the column, and then a linear gradient of 0.25 M NaCl in 12.5 mM Bis-tris pH 6.0 buffer was applied. Total gradient volume 540 ml. 10 ml fractions were collected. The peroxidase eluted in 20 ml at 0.16 M NaCl. 4 ml were applied to a column of Sephadex G 25 SF (16 mm Ø 400 mm) packed in 0.1 M sodium phosphate pH 7 at a flow rate of 4 ml/min. 2 ml fractions were collected. The peroxidase was eluted in 6 ml.

Conditions for testing hydrogen peroxide stability:

Enzyme	25 nM
Hydrogen peroxide	0.2 mM
Temperature	30°C
pH	10.5
Carbonate buffer	20 mM

Reagents:

Sodium carbonate buffer 22 mM pH 10.5: 1.807 g of Na_2CO_3 and 0.416 g of NaHCO_3 are dissolved in deionised water to a final volume of 1 l. pH is checked.

H_2O_2 : 2.2 mM.

Peroxidase solution: The pool from Sephadex G25 SF is diluted with sodium carbonate buffer 22 mM pH 10.5 to $\text{OD}_{404} = 0.0030$. (This corresponds to 27.5 nM).

Accomplishment:

The sodium carbonate buffer is preheated to 30°C. The peroxidase solution is made. A sample of the peroxidase solution is immediately withdrawn and diluted and submitted to the ABTS-assay. 5 ml of the peroxidase solution are mixed with 0.5 ml of 2.2 mM H₂O₂ and the mixture is placed in a water bath at 30°C. After 4, 8, 12 and 16 min samples are withdrawn, diluted and submitted to the ABTS-assay. Residual activities are fitted to 1st order decay and half-lives are calculated. rCiP is always included.

The results of the hydrogen peroxide stability test are presented in Table 1 below.

Table 1

Variant	T _½ (min)	Index
rCiP	6.5	1.00
E239K	8.3	1.27
Y272F	8.7	1.34
M276S	7.3	1.13
M242I + Y272F + E239K	14.8	2.27

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT:

- 5 (A) NAME: NOVO NORDISK A/S, NN
(B) STREET: Novo Alle
(C) CITY: Bagsvaerd
(E) COUNTRY: Denmark
(F) POSTAL CODE (ZIP): DK-2880
(G) TELEPHONE: +45 44 44 88 88
10 (H) TELEFAX: +45 44 49 05 55
(I) TELEX: 37173

(ii) TITLE OF INVENTION: PEROXIDASE VARIANTS 2

(iii) NUMBER OF SEQUENCES: 1

15 (iv) COMPUTER READABLE FORM:

- (A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.25 (EPO)

20 (v) CURRENT APPLICATION DATA:
APPLICATION NUMBER:

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

- 25 (A) LENGTH: 343 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vi) ORIGINAL SOURCE:

- 30 (A) ORGANISM: Coprinus cinereus
(B) STRAIN: IFO 8371

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

Gln Gly Pro Gly Gly Gly Gly Ser Val Thr Cys Pro Gly Gly Gln Ser
1 5 10 15
35 Thr Ser Asn Ser Gln Cys Cys Val Trp Phe Asp Val Leu Asp Asp Leu
20 25 30
Gln Thr Asn Phe Tyr Gln Gly Ser Lys Cys Glu Ser Pro Val Arg Lys
35 40 45
40 Ile Leu Arg Ile Val Phe His Asp Ala Ile Gly Phe Ser Pro Ala Leu
50 55 60

Thr Ala Ala Gly Gln Phe Gly Gly Gly Gly Ala Asp Gly Ser Ile Ile
 65 70 75 80
 Ala His Ser Asn Ile Glu Leu Ala Phe Pro Ala Asn Gly Gly Leu Thr
 85 90 95
 5 Asp Thr Val Glu Ala Leu Arg Ala Val Gly Ile Asn His Gly Val Ser
 100 105 110
 Phe Gly Asp Leu Ile Gln Phe Ala Thr Ala Val Gly Met Ser Asn Cys
 115 120 125
 10 Pro Gly Ser Pro Arg Leu Glu Phe Leu Thr Gly Arg Ser Asn Ser Ser
 130 135 140
 Gln Pro Ser Pro Pro Ser Leu Ile Pro Gly Pro Gly Asn Thr Val Thr
 145 150 155 160
 Ala Ile Leu Asp Arg Met Gly Asp Ala Gly Phe Ser Pro Asp Glu Val
 165 170 175
 15 Val Asp Leu Leu Ala Ala His Ser Leu Ala Ser Gln Glu Gly Leu Asn
 180 185 190
 Ser Ala Ile Phe Arg Ser Pro Leu Asp Ser Thr Pro Gln Val Phe Asp
 195 200 205
 20 Thr Gln Phe Tyr Ile Glu Thr Leu Leu Lys Gly Thr Thr Gln Pro Gly
 210 215 220
 Pro Ser Leu Gly Phe Ala Glu Glu Leu Ser Pro Phe Pro Gly Glu Phe
 225 230 235 240
 Arg Met Arg Ser Asp Ala Leu Leu Ala Arg Asp Ser Arg Thr Ala Cys
 245 250 255
 25 Arg Trp Gln Ser Met Thr Ser Ser Asn Glu Val Met Gly Gln Arg Tyr
 260 265 270
 Arg Ala Ala Met Ala Lys Met Ser Val Leu Gly Phe Asp Arg Asn Ala
 275 280 285
 30 Leu Thr Asp Cys Ser Asp Val Ile Pro Ser Ala Val Ser Asn Asn Ala
 290 295 300
 Ala Pro Val Ile Pro Gly Gly Leu Thr Val Asp Asp Ile Glu Val Ser
 305 310 315 320
 Cys Pro Ser Glu Pro Phe Pro Glu Ile Ala Thr Ala Ser Gly Pro Leu
 325 330 335
 35 Pro Ser Leu Ala Pro Ala Pro
 340

CLAIMS

1. A peroxidase variant with improved hydrogen peroxide stability at alkaline conditions, characterized by insertion, deletion or substitution of one or more amino acid residues in the region from amino acid residue 48 to 56, 76, 109, 214, 239, 258 to 262, 264, 266 to 272 of the parent peroxidase, a Coprinus cinereus peroxidase encoded by the amino acid sequence shown in SEQ ID No. 1.

2. A peroxidase variant according to claim 1, wherein one or more amino acid residues are substituted as follows

K48S,
V53K,A
H109K,
E214L,
15 E239A,V,L,I,P,F,W,M,G,S,T,C,Y,N,Q,D,K,R,H,
in particular E239K,G,S,L,Q,M,
W258F,H,
M261A,V,L,I,P,F,W,G,S,T,C,Y,N,Q,D,E,K,R,H,
in particular M261F,V,I,L,Q,
20 M268A,V,L,I,P,F,W,G,S,T,C,Y,N,Q,D,E,K,R,H,
in particular M268F,V,I,L,Q,
Y272A,V,L,I,P,F,W,G,S,T,C,M,N,Q,D,E,K,R,H,
in particular Y272F.

3. A peroxidase variant with improved hydrogen peroxide stability at alkaline conditions, wherein one or more amino acid residues of the parent peroxidase, a Coprinus cinereus peroxidase encoded by the amino acid sequence shown in SEQ ID No. 1, are substituted as follows

G72Q,
30 A91C,
N92K,
Q118E,
M125A,P,W,G,S,T,C,Y,N,D,E,K,R,H,
in particular M125S,T,

S147Q,
I152C,
P155C,
M166A, P, W, G, S, T, C, Y, N, D, E, K, R, H,
5 in particular M166S, T,
N192K,
I195K,
V206R,
K218R,
10 F229G,
A230C,
M242A, P, W, G, S, T, C, Y, N, D, E, K, R, H,
in particular M242S, T,
S244C,
15 S252P,
M276A, P, W, G, S, T, C, Y, N, D, E, K, R, H,
in particular M276S, T,
K278R,
M279A, P, W, G, S, T, C, Y, N, D, E, K, R, H,
20 in particular M279S, T,
A304E,
V314P,
K41R + K48R,
V53K + Q118E,
25 V53A + E239G,
I152C + A91C,
P155C + A230C,
M166F + E239K,
G167N + V176L,
30 E214L + E239L,
R241E + E239K,
S244C + P155C,
E239K + M242I + Y272F,
M166F + E239K + M242I + Y272F,
35 M125L + M166F + E239K + M242I + Y272F.

4. A peroxidase variant which is a fragment of a

peroxidase variant according to any of claims 1-3.

5. A bleaching composition comprising a peroxidase variant according to any of claim 1-4 and hydrogen peroxide or a hydrogen peroxide precursor, e.g. perborate or percarbonate, or a hydrogen peroxide generating enzyme system, e.g. an oxidase and its substrate, or a peroxy-carboxylic acid or a salt thereof.

6. A bleaching composition according to claim 5, wherein the amount of peroxidase variant corresponds to a concentration in the wash liquor of between 0.01 and 20 PODU/ml, and the amount of hydrogen peroxide or hydrogen peroxide precursor or hydrogen peroxide generating enzyme system or peroxy-carboxylic acid or a salt thereof corresponds to a hydrogen peroxide concentration in the wash liquor of up to 20 mM H_2O_2 .

7. A bleaching composition according to claim 5 or 6, which additionally comprises an oxidizable substrate such as an organic compound, such as a phenolic compound or a derivative of a phenothiazine or a derivative of a phenoxazine.

8. A bleaching composition according to claim 7, wherein the amount of oxidizable substrate corresponds to a concentration in the wash liquor of between 0.1 μM and 100 μM .

9. A detergent composition comprising a surfactant, a peroxidase variant according to any of claims 1-4 and hydrogen peroxide or a hydrogen peroxide precursor, e.g. perborate or percarbonate, or a hydrogen peroxide generating enzyme system, e.g. an oxidase and its substrate, or a peroxy-carboxylic acid or a salt thereof.

10. A detergent composition according to claim 9, wherein the amount of peroxidase variant corresponds to a concentration in the wash liquor of between 0.01 to 20 PODU/ml,

and the amount of hydrogen peroxide or hydrogen peroxide precursor or hydrogen peroxide generating enzyme system or peroxy-carboxylic acid or a salt thereof corresponds to a hydrogen peroxide concentration in the wash liquor of up to 20 mM H_2O_2 .

11. A detergent composition according to claim 9 or 10, which additionally comprises an oxidizable substrate such as an organic compound, such as a phenolic compound, or a derivative of a phenothiazine or a derivative of a phenoxazine.

10 12. A detergent composition according to claim 11, wherein the amount of oxidizable substrate corresponds to a concentration in the wash liquor of between 0.1 μM and 100 μM .

1/1

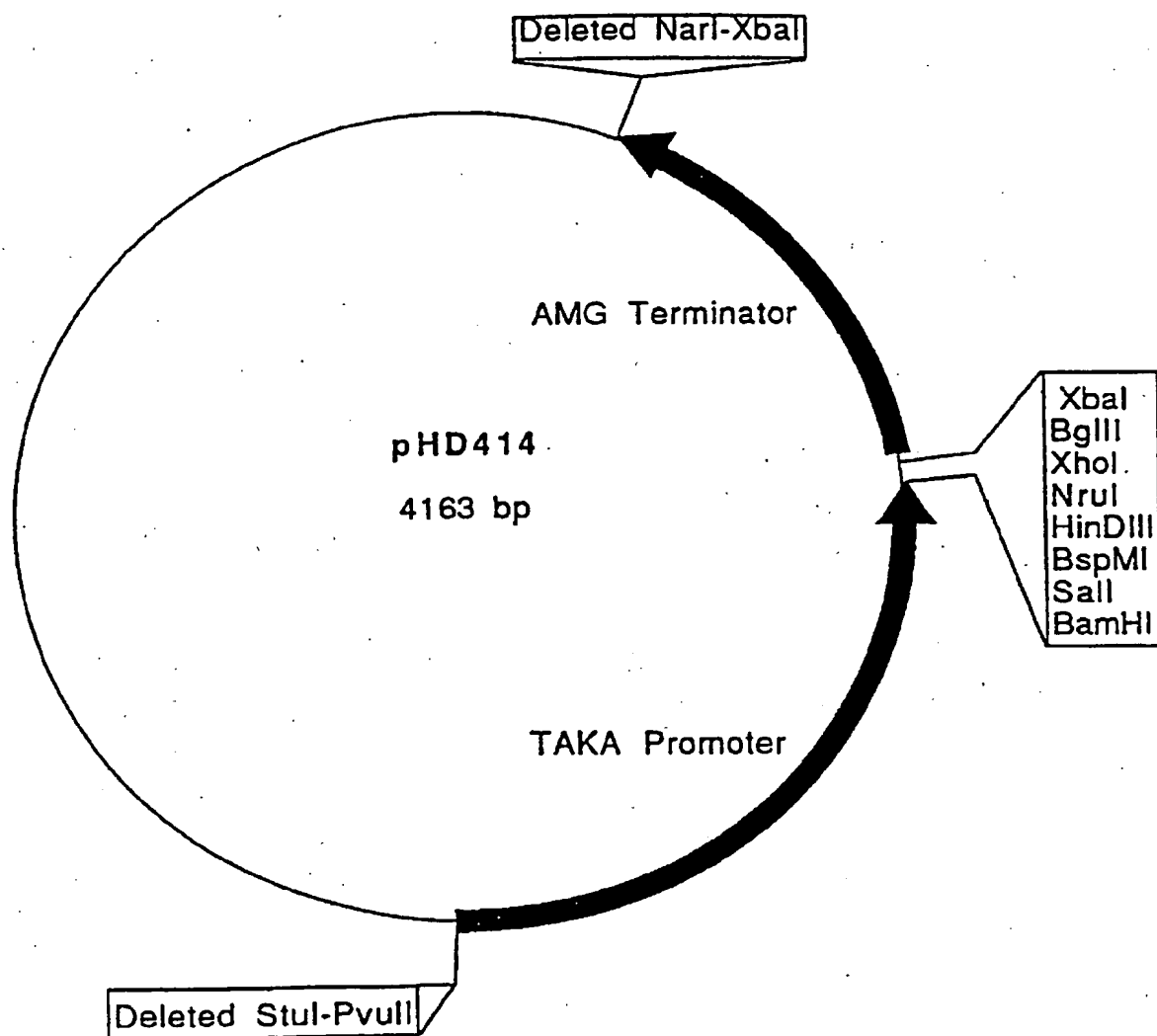


Fig. 1

1
INTERNATIONAL SEARCH REPORT

International application No.
PCT/DK 94/00382

A. CLASSIFICATION OF SUBJECT MATTER

IPC6: C12N 9/08, C11D 3/386

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC6: C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

SE,DK,FI,NO classes as above

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

BIOSIS, CA, WPI, DERWENT BIOTECHNOLOGY ABSTRACTS

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P,X	WO, A1, 9324618 (NOVO NORDISK A/S), 9 December 1993 (09.12.93), the whole document, see especially page 5 line 27 --	1-12
X	WO, A1, 9105858 (NOVO NORDISK A/S), 2 May 1991 (02.05.91), page 1, line 23 - line 25; page 3, line 1 - line 2; page 3, line 10 - line 26, page 6, line 18 - line 22 --	1-12
X	WO, A1, 9216634 (NOVO NORDISK A/S), 1 October 1992 (01.10.92), page 6, line 24 - page 7, line 11 --	1-4

☒ Further documents are listed in the continuation of Box C.

☒ See patent family annex.

* Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

"B" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance: the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance: the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search

16 January 1995

Date of mailing of the international search report

26 -01- 1995

Name and mailing address of the ISA/
Swedish Patent Office

Box 5055, S-102 42 STOCKHOLM

Facsimile No. +46 8 666 02 86

Authorized officer

Hans Bäckström

Telephone No. +46 8 782 25 00

INTERNATIONAL SEARCH REPORT

International application No.

PCT/DK 94/00382

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	EP, A1, 0299682 (AMERSHAM INTERNATIONAL PLC), 18 January 1989 (18.01.89), page 2, line 32 - line 36 --	1-4
A	EP, A2, 0179486 (SUNTORY LIMITED), 30 April 1986 (30.04.86) -----	1-12

INTERNATIONAL SEARCH REPORT

International application No.
PCT/DK 94/00382

Box I Observations where certain claims were found unsearchable (Continuation of Item 1 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

2. ☐ Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of Item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

See extra sheet

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☒ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:

4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
☐ No protest accompanied the payment of additional search fees.

According to rule 13.2, an international application shall relate to one invention only or a group of inventions linked by one or more of the same or corresponding "special technical features", i.e. features that define a contribution which each of the inventions makes over prior art.

Such a link between all the subject of claims 1-12 would be a peroxidase modified to improved stability in the presence of hydrogen peroxide. This *a priori* allegation however is not acceptable due to the state of the art as revealed in the attached search report, e.g. WO 92/16634 (p. 6 line 36 - p. 7 line 11) or WO 91/05858 (p. 3 line 10 - line 26).

Accordingly, the following inventions were found:

Invention 1, claims 1-12 partially: A peroxidase modified at any of amino acid residue 48-56 of the parent peroxidase enzyme (SEQ ID No. 1), a bleaching composition and a detergent composition comprising the peroxidase.

Invention 2, claims 1-12 partially: A peroxidase modified at amino acid residue 76 of the parent peroxidase enzyme (SEQ ID No. 1), a bleaching composition and a detergent composition comprising the peroxidase.

Invention 3, claims 1-12 partially: A peroxidase modified at amino acid residue 109 of the parent peroxidase enzyme (SEQ ID No. 1), a bleaching composition and a detergent composition comprising the peroxidase.

Invention 4, claims 1-12 partially: A peroxidase modified at amino acid residue 214 of the parent peroxidase enzyme (SEQ ID No. 1), a bleaching composition and a detergent composition comprising the peroxidase.

Invention 5, claims 1-12 partially: A peroxidase modified at any of amino acid residue 239 of the parent peroxidase enzyme (SEQ ID No. 1), a bleaching composition and a detergent composition comprising the peroxidase.

Invention 6, claims 1-12 partially: A peroxidase modified at any of amino acid residue 258-272 of the parent peroxidase enzyme (SEQ ID No. 1), a bleaching composition and a detergent composition comprising the peroxidase.

INTERNATIONAL SEARCH REPORT
Information on patent family members

31/12/94

International application No.
PCT/DK 94/00382

Patent document cited in search report		Publication date	Patent family member(s)		Publication date
WO-A1-	9324618	09/12/93	NONE		
WO-A1-	9105858	02/05/91	AT-T-	108484	15/07/94
			AU-B-	646645	03/03/94
			AU-A-	6515790	16/05/91
			AU-A-	6516090	16/05/91
			CA-A-	2067748	14/04/91
			CN-A-	1051600	22/05/91
			DE-D-	69010691	00/00/00
			EP-A,B-	0495836	29/07/92
			SE-T3-	0495836	
			EP-A,B-	0497794	12/08/92
			SE-T3-	0497794	
			ES-T-	2057593	16/10/94
			JP-T-	5500899	25/02/93
			JP-T-	5503542	10/06/93
			US-A-	5273896	28/12/93
			WO-A-	9105839	02/05/91
WO-A1-	9216634	01/10/92	BR-A-	9205802	28/06/94
			EP-A-	0505311	23/09/92
			JP-T-	6506108	14/07/94
EP-A1-	0299682	18/01/89	NONE		
EP-A2-	0179486	30/04/86	SE-T3-	0179486	
			DE-A-	3585133	20/02/92
			JP-C-	1642011	18/02/92
			JP-B-	3001949	11/01/91
			JP-A-	61104784	23/05/86
			US-A-	5116751	26/05/92